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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
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EXAMINER

GOLDBERG, JEANINE ANNE

ART UNIT PAPER NUMBER

1634

DATE MAILED: 07/08/2003

Please find below and/or attached an Office communication concerning this application or proceeding.

Office Action Summary	Application No.	Applicant(s)	
	09/601,645	DAHME ET AL.	
	Examiner	Art Unit	
	Jeanine A Goldberg	1634	

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --
Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If the period for reply specified above is less than thirty (30) days, a reply within the statutory minimum of thirty (30) days will be considered timely.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133).
- Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 06 May 2003.
- 2a) ☒ This action is **FINAL**. 2b) ☐ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-19,21-24,26-38,52-67 and 69-72 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-19,21-24,26-38,52-67 and 69-72 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
- 11) ☐ The proposed drawing correction filed on _____ is: a) ☐ approved b) ☐ disapproved by the Examiner.
If approved, corrected drawings are required in reply to this Office action.
- 12) ☐ The oath or declaration is objected to by the Examiner.

Priority under 35 U.S.C. §§ 119 and 120

- 13) ☒ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
2. ☐ Certified copies of the priority documents have been received in Application No. _____.
3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).
* See the attached detailed Office action for a list of the certified copies not received.
- 14) ☐ Acknowledgment is made of a claim for domestic priority under 35 U.S.C. § 119(e) (to a provisional application).
a) ☐ The translation of the foreign language provisional application has been received.
- 15) ☐ Acknowledgment is made of a claim for domestic priority under 35 U.S.C. §§ 120 and/or 121.

Attachment(s)

- | | |
|--|---|
| 1) <input type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413) Paper No(s). _____ |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | 5) <input type="checkbox"/> Notice of Informal Patent Application (PTO-152) |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO-1449) Paper No(s) _____ | 6) <input type="checkbox"/> Other: _____ |

DETAILED ACTION

1. This action is in response to the papers filed February 25, 2002; January 9, 2002. Currently, claims 1-19, 21-24, 26-38, 52-67, 69-72 are pending.
2. This action is FINAL.
3. Any objections and rejections not reiterated below are hereby withdrawn in view of the amendments to the claims and applicant's arguments.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

4. This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

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5. Newly amended Claims 1, 2, 4, 7-11, 14-19, 21-24, 26-28, 34-37, 38, 52-56, 60-64, 67, 69-70 and Newly added Claims 71-72 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) in view of Van Vlasselaer et al (US Pat. 5,648,223, July 1997).

It is noted that only the specification for the patent has been provided because the sequences are not relied upon in the rejection and are very extensive in length.

Cech et al. (herein referred to as Cech) teaches methods of quantitating tumor cells in a body fluid by concentrating tumor cells in a sample of body fluid, amplifying mRNA coding for the catalytic subunit of telomerase and quantitatively determining the amount of amplified nucleic acid. Specifically, Cech teaches methods of diagnosing cancer in a patient by obtaining a biological sample from the patient and detecting a hTERT gene product in the patient sample, where the detection of the hTERT gene product in the sample is correlated with a diagnosis of cancer (col. 6, lines 20-40).

Cech also teaches that the determination of an hTERT gene, mRNA or protein level above normal or standard range is indicative of the presence of telomerase-positive cells, or immortal of which certain tumor cells are examples (col. 99, lines 5-20). Cech specifically teaches that hTERT gene or gene product (i.e., mRNA or polypeptide) is preferably detected and/or quantified in a biological sample (col. 104, lines 59-65).

Cech teaches that biological samples include blood, blood cells, body fluids, e.g., urine, sputum, amniotic fluid, blood, peritoneal fluid, pleural fluid, semen (col. 104, lines 65-68)(limitations of Claim 11, 24-28). Cech teaches that cells or tissues may be fractionated before analysis, for example by a cell sorter may be used to sort cells

according to characteristics such as expression of a surface antigen (col. 105, lines 10-12). Cech teaches that nucleic acids may be isolated from the cell by any separation of the species or target to be detected from any other substance in the mixture (col. 105, lines 55-65)(limitations of Claim 4). Cech teaches that the assay for detection of hTERT are amplification based to amplify all or part of an hTERT gene or transcript where the amplification product is then detected directly or indirectly (col. 106, lines 45-55). Cech teaches primers useful for PCR amplification of hTERT are provided in Table 2 (col. 107, lines 1-5). Cech teaches that amplified products may be directly analyzed by size (gel electrophoresis); by hybridization to a target nucleic acid immobilized on a solid support; by sequencing; by detection of a fluorescent, phosphorescent, or radioactive signal (col. 108, lines 1-5)(limitations of Claim 8-9). Cech teaches that in one possible embodiment PCR amplification is carried out in a 50ul solution containing the nucleic acid sample (e.g., cDNA obtained through reverse transcription of hTERT RNA), dNTP, hTERT specific PCR primers, Taq polymerase, PCR buffer (col. 107, lines 12-25)(limitations of Claim 2, 14, 15). Cech teaches that quantification methods may include the co-amplification reactions to allow for normalization of the cell number in a sample as compared to the amount of hTERT in the sample (col. 108, lines 45-65)(limitations of Claim 7, 10).

Cech does not specifically teach enrichment by centrifugation to collect tumor cells.

However, Van Vlasselaer teaches methods for enriching tumor cells prior to analyzing. Percoll and Ficoll were routinely used in the art as cell separation media (col. 4, lines 60- col. 5, lines 5)(limitations of claim 23). Van Vlassalaer teaches that the

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medium density is adjusted to the density of the cell type (col. 9, lines 47-66, col. 14, Example 6.1.1 and 6.1.2)(limitations of Claim 21, 22, 24 and 62-64). Van Vlassalaer teaches that a large volume of complete blood may be directly placed on the density gradient. Peripheral blood may be collected in anti-coagulant containing tubes (col. 4, lines 37-40)(limitations of Claim 24-27). Van Vlassalaer teaches that for breast tumor cells, the specific density was adjusted within 0.0005 g/ml of the specific density of the tumor cells and the centrifugation speed is at a gravitational force sufficient to pellet the cells (col. 16, lines 61-67). Additionally, Van Vlasselaer teaches methods that may cause cells to be heavier than their normal density so that they are pelleted during centrifugation, namely linking a heavy particle such as a binding agent to selected cells. Van Vlassalaer teaches that centrifugation is carried out in a tube divided by a barrier wherein the barrier is an annular ring (col. 5, lines 30-54).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech with the enrichment method of Van Vlasselaer prior to molecular analysis. Van Vlasselaer teaches that after centrifugation is performed and cells are collected, tumor cell may be screened by molecular means (col. 4 lines 20-22). Therefore, the ordinary artisan would have been motivated to have performed the rapid and high yield isolation or enrichment of tumor cells prior to analysis by the method of Cech for the increased sensitivity and efficiency. The numerous means of enrichment and isolation of Van Vlasselaer would be obvious means to enriching cells in a blood sample. Therefore, it would have been obvious to

the ordinary artisan to employ a cell separation medium that was proven in the art and readily available.

With regard to Claims 21, 22, 24, 62-64, it would have been obvious to the ordinary artisan to adjust the density of the cell separation medium and centrifugation speed according to the type of tumor cell to be concentrated. Van Vlasselaer specifically teaches that methods for determining the specific density of a given tumor cell is described *infra* (col. 4, lines 10-15). Additionally, Van Vlasselaer teaches methods that may cause cells to be heavier than their normal density so that they are pelleted during centrifugation, namely linking a heavy particle such as a binding agent to selected cells. Therefore, the determination of cell densities requires routine experimentation. With regard to the densities of certain cells, these are routinely optimizable based upon the desired parameters, since Van Vlasselaer teaches how densities may be determined the optimization of the workable density is not inventive. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation." It would have been obvious to provide a substance that prevents platelets from sticking to the tumor cells and to remove the platelets as routinely practiced in the art.

Response to Arguments

The response traverses the rejection. The response asserts that all of the claims include one element, namely the method for the quantification of tumor cells in a body fluid. The response argues that neither of the cited references teaches or suggests a

method for the quantification of tumor cells in a body fluid. This argument has been reviewed but is not convincing because Cech teaches quantifying the amount of amplified nucleic acid. Therefore, Cech teaches each of the positive process steps of the method. Moreover, the response argues that Cech does not teach or suggest correlating the amount of the amplified mRNA to the number of tumor cells in the sample. The claims, as written, state, "quantitatively determining the amount of amplified nucleic acid, thereby quantifying tumor cells in a body fluid." Thus, the claim clearly implies that by merely determining the nucleic acid, the tumor cells have been quantified. The language "thereby" is not a positive process step. Additionally, the language implies that no additional steps are required to determine tumor cells since the quantification of amplified nucleic acid "thereby" quantifies tumor cells.

The response argues that Van Vlasselaer does not teach any diagnostic assay for the detection or quantification of tumor cells in general in a body fluid. This argument has been thoroughly reviewed, but is not found persuasive because Van Vlasselaer specifically teaches how to determine the "specific density of a given tumor cell." (col. 4, lines 5-15). Once a density range is determined where a given tumor cell is found, fine tuning of the density gradient can be performed (col. 4, lines 16-20). Therefore, the teachings of Van Vlasselaer clearly demonstrates how to determine and enrich tumor cells using density gradients. While Van Vlasselaer exemplifies breast tumor cells, the teachings of Van Vlasselaer are broader.

The response asserts that the combination of references fails to teach the method as a whole. This argument has been thoroughly reviewed, but is not found

persuasive because given the specific teachings in Cech that cells or tissues may be fractionated before analysis and the clear teachings of Van Vlasselaer of how to fractionate tumor cells, the skilled artisan would be motivated to separate tumor cells using the method of Van Vlasselaer prior to analysis of Cech.

With respect to Claim 71, the ordinary artisan would have been motivated to isolate telomerase-positive non-tumor cells from tumor cells to eliminate background within the assay. Given the teachings of Van Vlasselaer, the skilled artisan would have a reasonable expectation of success for separating various cells based upon density. Van Vlaselaer provides specific methods for determining the specific density of a given tumor cell.

The response asserts that Judicial notice is not appropriate in the instant case. The response objects to items routinely practiced in the art. For example, the response asserts that the examiner can not take official notice of facts outside the record, such as that Percoll and Ficoll as cell separation media. It is noted that the examiner has not taken judicial notice with respect to Percoll and Ficoll. Van Vlasselaer, as specifically pointed out in the office action, page 7, teaches Percoll and Ficoll was used in the art as cell separation media (col. 4, lines 60- col. 5, lines 5)(limitations of Claim 23). Therefore, the examiner has not taken judicial notice on this limitation as asserted by the response.

The response further asserts that judicial notice was taken for "providing a substance that prevents platelets from sticking to the tumor cells and facilitates removal of the platelets." Again, the examiner has specifically pointed to a cite in Van Vlasselaer

which teaches this limitation. Peripheral blood may be collected in anti-coagulant containing tubes (col. 4, lines 37-40)(limitations of Claim 24-27). Therefore, the examiner has not taken judicial notice for this limitation.

Finally, the response argues that adjusting the density of the cell separation medium according to cell type is inappropriate judicial notice. It is noted that Van Vlasselaer states, "the medium density is adjusted to the density of the cell type (col. 9, lines 47-66, col. 14, Example 6.1.1 and 6.1.2)(limitations of Claim 21, 22, 24 and 62-64). The response may have mistaken routine optimization for judicial notice. However, routine optimization is not the same as judicial notice. The reference specifically teaches that specific densities may be ascertained, and the skilled artisan would be within the skill of the art to determine various densities. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation." Thus, the optimum or workable densities of specific cells is routine experimentation.

Thus for the reasons above and those already of record, the rejection is maintained.

6. Claim 3 is rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Gwynn et al. (US Pat. 6,025,156, February 2000).

Cech does not specifically teach using DNAase for removal of DNA from a sample.

However, Gwynn et al. (herein referred to as Gwynn) teaches using DNAase for removal of DNA from RNA samples. Once the DNAase was added and DNA was removed, RNA was pelleted and reverse transcribed into cDNA (col. 31, lines 50-65).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of generating cDNA of Cech with the teachings of Gwynn. Gwynn teaches addition of DNAase facilitates the removal of DNA from the sample such that RNA may be obtained. Therefore, the ordinary artisan would have realized that in order to remove DNA from RNA samples so that the RNA may be in turn transcribed, DNAase may be added.

Response to Arguments

The response traverses the rejection. The response asserts that all of the claims include one element, namely the method for the quantification of tumor cells in a body fluid. The response argues that neither of the cited references teaches or suggests a method for the quantification of tumor cells in a body fluid. This argument has been reviewed but is not convincing because Cech teaches quantifying the amount of amplified nucleic acid. Therefore, Cech teaches each of the positive process steps of the method. Moreover, the response argues that Cech does not teach or suggest correlating the amount of the amplified mRNA to the number of tumor cells in the sample. The claims, as written, state, "quantitatively determining the amount of amplified nucleic acid, thereby quantifying tumor cells in a body fluid." Thus, the claim

clearly implies that by merely determining the nucleic acid, the tumor cells have been quantified. The language "thereby" is not a positive process step. Additionally, the language implies that no additional steps are required to determine tumor cells since the quantification of amplified nucleic acid "thereby" quantifies tumor cells.

In response to applicant's argument that Gwynn is nonanalogous art, it has been held that a prior art reference must either be in the field of applicant's endeavor or, if not, then be reasonably pertinent to the particular problem with which the applicant was concerned, in order to be relied upon as a basis for rejection of the claimed invention. See *In re Oetiker*, 977 F.2d 1443, 24 USPQ2d 1443 (Fed. Cir. 1992). In this case, Gwynn is within the same field of applicant's endeavor and is directed to purification of nucleic acids. The elimination of DNA would provide a more pure RNA sample.

Gwynn has not been used to suggest reverse transcribing and specifically amplifying the catalytic subunit of telomerase. The primary reference, Cech has been relied upon to provide the teachings.

Thus for the reasons above and those already of record, the rejection is maintained.

7. Claims 29 is rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Shelby (GB 2 260 811, April 1993).

Cech does not specifically teach a method of centrifugation by the allowing of the sample to cool following centrifugation.

However, Shelby teaches that diagnosis or monitoring of cancer or a malignant tumor may be effected by the detection of mRNA in a sample such as peripheral blood where the cells are not normally present and testing the sample (abstract). The detection technique of Shelby involves extracting the total cellular mRNA in a sample using reverse transcriptase to prepare cDNA, then carrying out PCR with appropriate primers so as to selectively amplify the cDNA (abstract). Shelby teaches cooling after centrifugation was routinely practice in the art (page 10, last paragraph).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of centrifugation by the allowing of the sample to cool following centrifugation.

Response to Arguments

The response traverses the rejection. The response asserts that all of the claims include one element, namely the method for the quantification of tumor cells in a body fluid. The response argues that neither of the cited references teaches or suggests a method for the quantification of tumor cells in a body fluid. This argument has been reviewed but is not convincing because Cech teaches quantifying the amount of amplified nucleic acid. Therefore, Cech teaches each of the positive process steps of the method. Moreover, the response argues that Cech does not teach or suggest correlating the amount of the amplified mRNA to the number of tumor cells in the sample. The claims, as written, state, "quantitatively determining the amount of amplified nucleic acid, thereby quantifying tumor cells in a body fluid." Thus, the claim clearly implies that by merely determining the nucleic acid, the tumor cells have been

quantified. The language "thereby" is not a positive process step. Additionally, the language implies that no additional steps are required to determine tumor cells since the quantification of amplified nucleic acid "thereby" quantifies tumor cells.

Selby has not been used to suggest a method for concentrating or depleting tumor cells. The primary reference, Cech has been relied upon to provide the teachings. Selby is only relied upon for the purposes of indicating a cooling step is allowed. In response to applicant's arguments against the references individually, one cannot show nonobviousness by attacking references individually where the rejections are based on combinations of references. See *In re Keller*, 642 F.2d 413, 208 USPQ 871 (CCPA 1981); *In re Merck & Co.*, 800 F.2d 1091, 231 USPQ 375 (Fed. Cir. 1986).

The response asserts that Judicial notice is not appropriate in the instant case. The response objects to items routinely practiced in the art. For example, the response asserts that the examiner can not take official notice of facts outside the record, such as that cooling after centrifugation was routinely practiced in the art. It is noted that the examiner has not taken judicial notice with respect to cooling following centrifugation. Selby, as specifically pointed out in the office action, page 10, teaches cooling after centrifugation was routinely practice in the art (page 10, last paragraph). Therefore, the examiner has not taken judicial notice on this limitation as asserted by the response.

Thus for the reasons above and those already of record, the rejection is maintained.

8. Claims 5-6 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Gelmini et al. (Clinical Chemistry, Vol. 43, No. 5, pages 752-758, 1997).

Cech does not specifically teach the continuous monitoring of PCR reaction.

However, Gelmini teaches methods of quantitative polymerase chain reaction which is quantitative, accurate, and time-saving. The method of Gelmini uses fluorogenic probes to assess amplification. Gelmini teaches that during PCR cycling, the probe specifically hybridizes to the corresponding template and then is cleaved and results in increase of fluorescence emission of the reporter dye. The increased of fluorescence is proportional to the amount of the specific PCR product (page 753, col. 2). Gelmini teaches that the fluorescence was measured with a luminescence spectrometer (page 754, col. 1).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech for amplification with the method of Gelmini for the real time quantitative PCR. The ordinary artisan would have been motivated to have applied the method of Gelmini because Gelmini teaches the TaqMan PCR assay gave accurate, quantitative results.

Response to Arguments

The response traverses the rejection. The response asserts that all of the claims include one element, namely the method for the quantification of tumor cells in a body fluid. The response argues that neither of the cited references teaches or suggests a method for the quantification of tumor cells in a body fluid. This argument has been

reviewed but is not convincing because Cech teaches quantifying the amount of amplified nucleic acid. Therefore, Cech teaches each of the positive process steps of the method. Moreover, the response argues that Cech does not teach or suggest correlating the amount of the amplified mRNA to the number of tumor cells in the sample. The claims, as written, state, "quantitatively determining the amount of amplified nucleic acid, thereby quantifying tumor cells in a body fluid." Thus, the claim clearly implies that by merely determining the nucleic acid, the tumor cells have been quantified. The language "thereby" is not a positive process step. Additionally, the language implies that no additional steps are required to determine tumor cells since the quantification of amplified nucleic acid "thereby" quantifies tumor cells.

Gelmini has not been used to suggest a method for concentrating or depleting tumor cells. The primary reference, Cech has been relied upon to provide the teachings. Gelmini has only been relied upon to illustrate a quantitative detection PCR method. In response to applicant's arguments against the references individually, one cannot show nonobviousness by attacking references individually where the rejections are based on combinations of references. See *In re Keller*, 642 F.2d 413, 208 USPQ 871 (CCPA 1981); *In re Merck & Co.*, 800 F.2d 1091, 231 USPQ 375 (Fed. Cir. 1986).

Thus for the reasons above and those already of record, the rejection is maintained.

9. Claims 13 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997).

Cech does not specifically teach SEQ ID NO: 1 and 2 however, Cech teaches numerous primers suitable for PCR amplification of hTERT.

The nucleic acid sequences of SEQ ID NO: 1 and 2 are functional equivalents to the primers taught by Cech. The ordinary artisan would have recognized that primer designed to amplify all or part of an hTERT gene may be used. Cech teaches the parameters needed to design appropriate primers for hTERT. For example, Cech teaches that the primer are sufficiently complementary to the hTERT gene. The primers are typically at least 6 bases in length, typically between about 12 and about 50 bases (col. 106, lines 45-68). Cech teaches that one of skill in the art having the disclosure will be able, using routine methods will select primer to amplify all or any portion of hTERT gene. Therefore, SEQ ID NO: 1 and 2 of the instant application were merely selected by the routine methods provided by Cech for the amplification of all or part of the hTERT nucleic acid.

Response to Arguments

The response traverses the rejection. The response asserts that since Cech provides a laundry list of primers, there is no teaching or suggestion as to the selection of the particular sequences of primers which may be used. The response even asserts that the teachings of Cech teach away from the instant primers. This argument has been thoroughly reviewed, but is not found persuasive because the teachings of Cech provide clear guidance how to design primers which may be used to amplify the

catalytic subunit of telomerase. There is a reasonable expectation of success that the selection of primers within the guidelines presented by Cech would provide a primer which would amplify the nucleic acid. As discussed above, it was well known at the time of the invention how to design primers. Therefore, designing primers according to Cech would have been obvious and well within the skill of the artisan. Selecting SEQ ID NO: 1 and 2, are functional equivalents to the extensive list provided by Cech. It is evident from the extensive list in Cech that any number of primers would function to amplify the nucleic acid. Therefore, the primers would have been obvious to the skilled artisan at the time the invention was made.

The response asserts that Judicial notice is not appropriate in the instant case. The response objects to items routinely practiced in the art. For example, the response asserts that the examiner can not take official notice of facts outside the record, such as that cooling after centrifugation was routinely practiced in the art. It is noted that the examiner has not taken judicial notice with respect to the design of primers. The references specifically provides the guidelines for which how to design primers. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation." Routine optimization is not considered inventive and no evidence has been presented that the probe selection performed was other than routine, that the products resulting from the optimization have any unexpected properties, or that the results should be considered unexpected in any

way as compared to the closest prior art. Therefore, the examiner has not taken judicial notice on this limitation as asserted by the response.

Thus for the reasons above and those already of record, the rejection is maintained.

10. Claims 12, 57-59 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997) above in view of Melvin et al. (WO 97/12246, April 1997).

Cech does not specifically teach using controls.

However, Melvin et al. (herein referred to as Melvin) teaches that in RT-PCR experiments B-actin was used as a positive control. Melvin teaches that the negative control was sterile water in place of cDNA (page 17).

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech to include controls as taught by Melvin. Controls are essential in each scientific experiment to ensure that the results obtained are due to the experiment and not due to external factors. Therefore, the ordinary artisan would be motivated to have used controls in the study of Cech.

Response to Arguments

The response traverses the rejection. The response asserts that all of the claims include one element, namely the method for the quantification of tumor cells in a body fluid. The response argues that neither of the cited references teaches or suggests a method for the quantification of tumor cells in a body fluid. This argument has been

reviewed but is not convincing because Cech teaches quantifying the amount of amplified nucleic acid. Therefore, Cech teaches each of the positive process steps of the method. Moreover, the response argues that Cech does not teach or suggest correlating the amount of the amplified mRNA to the number of tumor cells in the sample. The claims, as written, state, "quantitatively determining the amount of amplified nucleic acid, thereby quantifying tumor cells in a body fluid." Thus, the claim clearly implies that by merely determining the nucleic acid, the tumor cells have been quantified. The language "thereby" is not a positive process step. Additionally, the language implies that no additional steps are required to determine tumor cells since the quantification of amplified nucleic acid "thereby" quantifies tumor cells.

Melvin has not been used to suggest a method for concentrating or depleting tumor cells. The primary reference, Cech has been relied upon to provide the teachings. Melvin has only been relied upon to illustrate the use of controls. In response to applicant's arguments against the references individually, one cannot show nonobviousness by attacking references individually where the rejections are based on combinations of references. See *In re Keller*, 642 F.2d 413, 208 USPQ 871 (CCPA 1981); *In re Merck & Co.*, 800 F.2d 1091, 231 USPQ 375 (Fed. Cir. 1986).

Thus for the reasons above and those already of record, the rejection is maintained.

11. Claims 30-33, 65-66 are rejected under 35 U.S.C. 103(a) as being unpatentable over by Cech et al (US Pat. 6,166,178, December 26, 2000, filed November 19, 1997)

in view of Van Vlasselaer et al (US Pat. 5,648,223, July 1997) as applied to Claims 18-28, 60-64 above, and further in view of Oka et al (US Pat. 5,298,165, March 1994).

Neither Cech nor Van Vlasselaer specifically teach centrifugation with filters of porous barriers which have certain properties.

However Oka et al. (herein referred to as Oka) teaches that filtration of blood may be effected with different membranes, filters or porous barriers. Oka teaches that the average pore size of one of the filters is preferably from 4 to 25 μm (col. 10, lines 40-45). Additionally, Oka teaches numerous different filters with different thickness including thickness of 2 mm, 5 mm.

Therefore, it would have been prima facie obvious to one of ordinary skill in the art to have modified the method of Cech in view of Van Vlasselaer to enrich using a porous barrier, filter or sieve as taught by Oka for the express benefit of enriching or isolating cells. The filters of Oka are representative of filters taught in the art. As exemplified by Oka the specific specifications of the filter are dependent upon the material wishing to be isolated. Therefore, with regard to the pore size and thickness of filters, these are routinely optimizable based upon the desired parameters, since Oka teaches how densities may be determined the optimization of the workable pore size and thickness is not inventive. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation."

Response to Arguments

The response traverses the rejection. The response asserts that all of the claims include one element, namely the method for the quantification of tumor cells in a body fluid. The response argues that neither of the cited references teaches or suggests a method for the quantification of tumor cells in a body fluid. This argument has been reviewed but is not convincing because Cech teaches quantifying the amount of amplified nucleic acid. Therefore, Cech teaches each of the positive process steps of the method. Moreover, the response argues that Cech does not teach or suggest correlating the amount of the amplified mRNA to the number of tumor cells in the sample. The claims, as written, state, "quantitatively determining the amount of amplified nucleic acid, thereby quantifying tumor cells in a body fluid." Thus, the claim clearly implies that by merely determining the nucleic acid, the tumor cells have been quantified. The language "thereby" is not a positive process step. Additionally, the language implies that no additional steps are required to determine tumor cells since the quantification of amplified nucleic acid "thereby" quantifies tumor cells.

Oka has not been used to suggest a method for concentrating or depleting tumor cells. The primary reference, Cech has been relied upon to provide the teachings. Oka has only been relied upon to illustrate teach centrifugation with filters of porous barriers which have certain properties. In response to applicant's arguments against the references individually, one cannot show nonobviousness by attacking references individually where the rejections are based on combinations of references. See *In re Keller*, 642 F.2d 413, 208 USPQ 871 (CCPA 1981); *In re Merck & Co.*, 800 F.2d 1091, 231 USPQ 375 (Fed. Cir. 1986).

Finally, the response argues that pore size and thicknesses of filters inappropriate judicial notice. Oka teaches that the average pore size of one of the filters is preferably from 4 to 25 um (col. 10, lines 40-45). Additionally, Oka teaches numerous different filters with different thickness including thickness of 2 mm, 5 mm. The response may have mistaken routine optimization for judicial notice. However, routine optimization is not the same as judicial notice. The reference specifically teaches that specific densities may be ascertained, and the skilled artisan would be within the skill of the art to determine various densities. As noted in *In re Aller*, 105 USPQ 233 at 235, "More particularly, where the general conditions of a claim are disclosed in the prior art, it is not inventive to discover the optimum or workable ranges by routine experimentation." Thus, the optimum or workable pore size of filters is not judicial notice.

Thus for the reasons above and those already of record, the rejection is maintained.

Conclusion

12. No claims allowable over the art.

13. Applicant's amendment necessitated the new ground(s) of rejection presented in this Office action. Accordingly, **THIS ACTION IS MADE FINAL**. See MPEP § 706.07(a). Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a).

A shortened statutory period for reply to this final action is set to expire **THREE MONTHS** from the mailing date of this action. In the event a first reply is filed within

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TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event, however, will the statutory period for reply expire later than SIX MONTHS from the date of this final action.

14. Any inquiry concerning this communication or earlier communications from the examiner should be directed to examiner Jeanine Goldberg whose telephone number is (703) 306-5817. The examiner can normally be reached Monday-Friday from 8:00 a.m. to 5:30 p.m.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Gary Jones, can be reached on (703) 308-1152. The fax number for this Group is (703) 305- 3014.

Any inquiry of a general nature should be directed to the Group receptionist whose telephone number is (703) 308-0196.


Jeanine Goldberg
July 1, 2003


B. J. FORMAN
PATENT EXAMINER